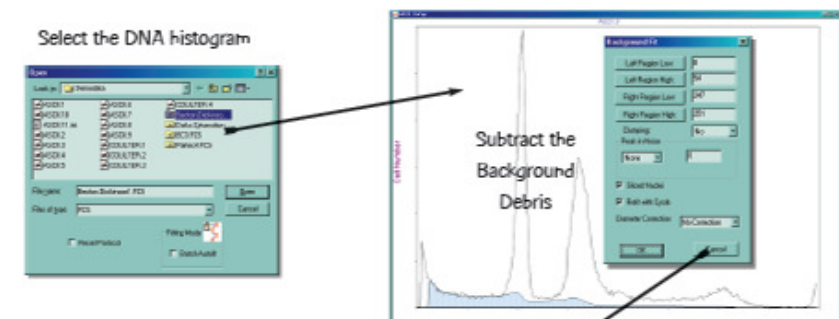


MultiCycle AV Standalone Analysis

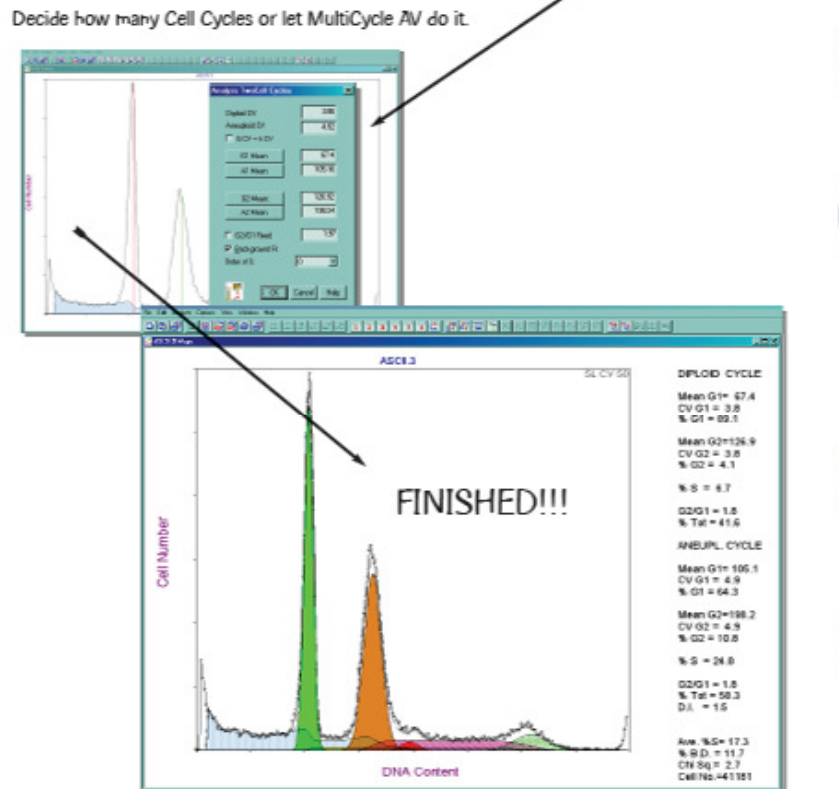
Included with the MultiCycle AV Plug-In for FCS Express™ is a standalone copy of MultiCycle AV at no extra charge

Select the DNA histogram



Subtract the Background Debris

Decide how many Cell Cycles or let MultiCycle AV do it.



FINISHED!!!

DNA Content

Cell Number

DIPLOID CYCLE

Mean G1	67.4
CV G1	3.5
% G1	52.5

ANEBURL CYCLE

Mean G1	105.1
CV G1	4.9
% G1	64.3

ANEBURL CYCLE

Mean G2	158.2
CV G2	4.9
% G2	10.0

ANEBURL CYCLE

Mean S	24.8
CV S	17.3
% S	11.7

ANEBURL CYCLE

Mean D	11.7
CV D	2.7
% D	41.151

For questions, pricing, ordering information please contact

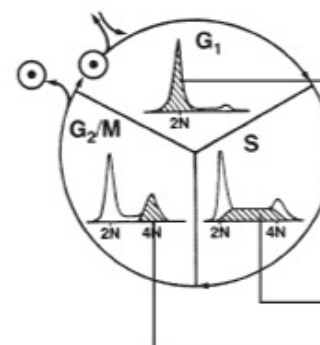


MultiCycle AV & FCS Express™

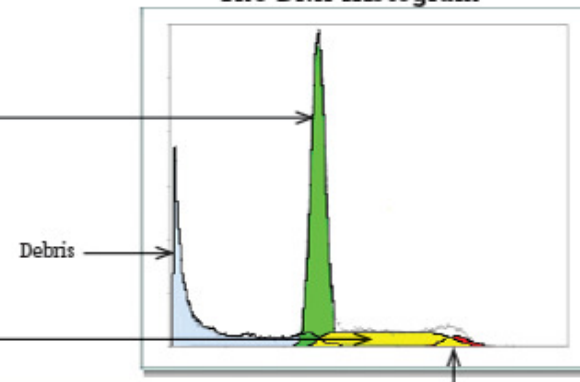
Powerful DNA Cell Cycle Analysis combined with Sophisticated List Mode Analysis

One of the earliest applications of flow cytometry was the measurement of DNA content in cells; the first technique available to characterize the non-mitotic phases of the cell cycle. This analysis is based on the ability of certain dyes to stain cellular DNA in a stoichiometric manner (the amount of stain is directly proportional to the amount of DNA within the cell).

The Biological Cell Cycle



The DNA Histogram



DNA content histograms require mathematical analysis in order to extract the underlying G₁, S, and G₂ phase distributions. Methods for this analysis have been developed and refined over the past two decades. The most flexible and accurate method of cell cycle analysis is based upon building a mathematical model of DNA content distribution, and then fitting this model to the data using curve-fitting methods. The most well established model, proposed by Dean and Jett (1974), is based upon the prediction that the cell cycle histogram is a result of the Gaussian broadening of the theoretically perfect distribution. The underlying distribution can be recovered or "deconvoluted" by fitting the G₁ and G₂ peaks as Gaussian curves and the S phase distribution as a Gaussian-broadened distribution.

Debris increases rapidly farther away. In the best case, the debris signal is an insignificant proportion of the histogram in the region occupied by the cell cycle data. Unfortunately this is often not the case in archived material, stored in paraffin, and accurate debris modeling is critically important in order to subtract the effects of the underlying debris from the cell cycle fitting. DNA analysis software that does not incorporate all the features of these models may yield accurate results in the ideal scenarios, but will often return incorrect results in even mildly complex situations. Worse, it is difficult to simply look at these results and appreciate that they are invalid.

Almost all cell or nuclear suspensions analyzed by DNA content flow cytometry contain some damaged or fragmented nuclei (debris) resulting in events, usually most visible to the left of the diploid G₁, which are not fit by the G₁, S or G₂ compartments. In samples that are derived from fresh tissues or cells, most of the "debris" is visible close to the origin of the histogram and de-

MultiCycle AV has a twenty year track record and incorporates several variations of the Dean and Jett model in order to accommodate a wide variety of experimental outcomes and cell types. In addition a "histogram dependent" exponential model or a "sliced-nuclei" model accurately removes the debris component from the true DNA histogram. More information about the justification and mathematics of these models are available at www.phoenixflow.com/cellcyclemath.pdf

